

# *Faecal indicator sensing – toward a faster method for marine & freshwater*

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# Outline

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- Drivers
- Background
- Results
- Rapid method-initial tests.
- Conclusions/Impact

# Drivers

## ➤ 2006/7/EC BWD

Inland waters					
	Parameter	Excellent quality	Good quality	Sufficient quality	Reference methods of analysis
1	Intestinal <i>Enterococci</i> (cfu/100mL)	200(*)	400(*)	330(*)	ISO 7899-1 or ISO 7899-2
2	<i>Escherichia coli</i> (cfu/100mL)	500(*)	1,000(*)	900(**)	ISO 9308-3 or ISO 9308-1

(\*) Based upon a 95-percentile evaluation, (\*\*) Based on a 90-percentile evaluation.

Coastal and transitional waters					
	Parameter	Excellent quality	Good quality	Sufficient quality	Reference methods of analysis
1	Intestinal <i>Enterococci</i> (cfu/100 mL)	100(*)	200(*)	185(*)	ISO 7899-1 or ISO 7899-2
2	<i>Escherichia coli</i> (cfu/100mL)	250(*)	500(*)	500(**)	ISO 9308-3 or ISO 9308-1

(\*) Based upon a 95-percentile evaluation, (\*\*) Based on a 90-percentile evaluation.

# Drivers

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- Public protection



# Standard Methods – *E. Coli*

Laborious

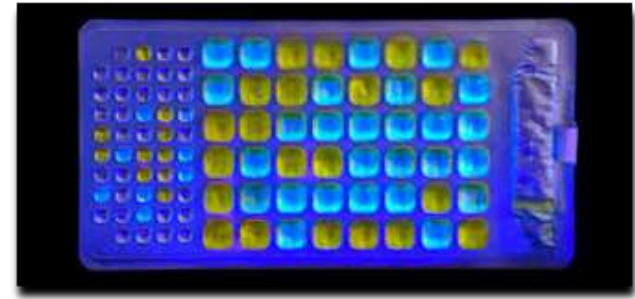
Not suitable when  
Immediate action  
required!

Expensive

Time  
Consuming  
(18-72 h)

# Enzyme based methods

$\beta$ -D-Glucuronidase  
(GUD)



Colilert 18

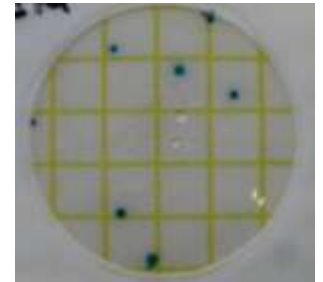
GUD extraction



GUD activity  
measurement



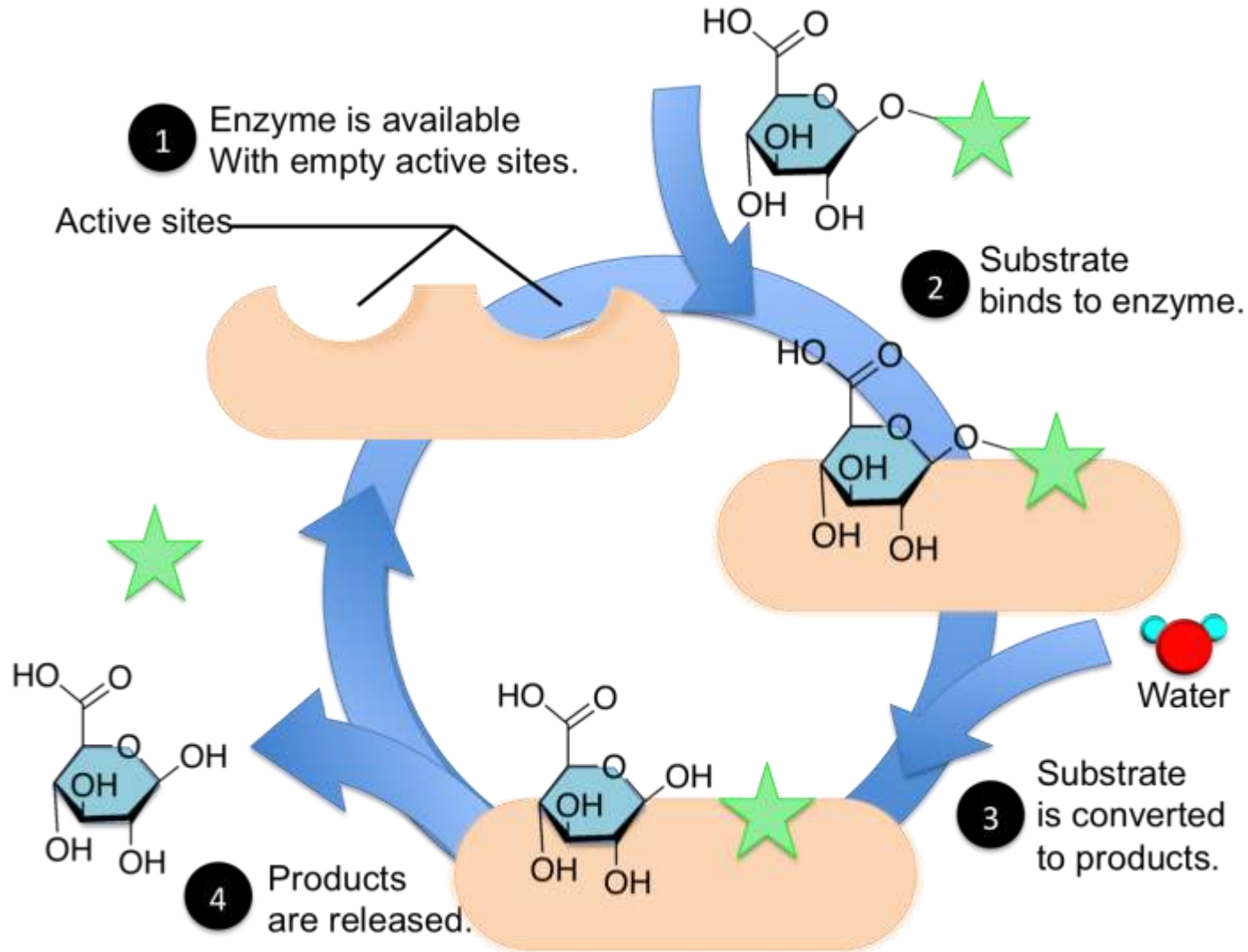
Chromocult



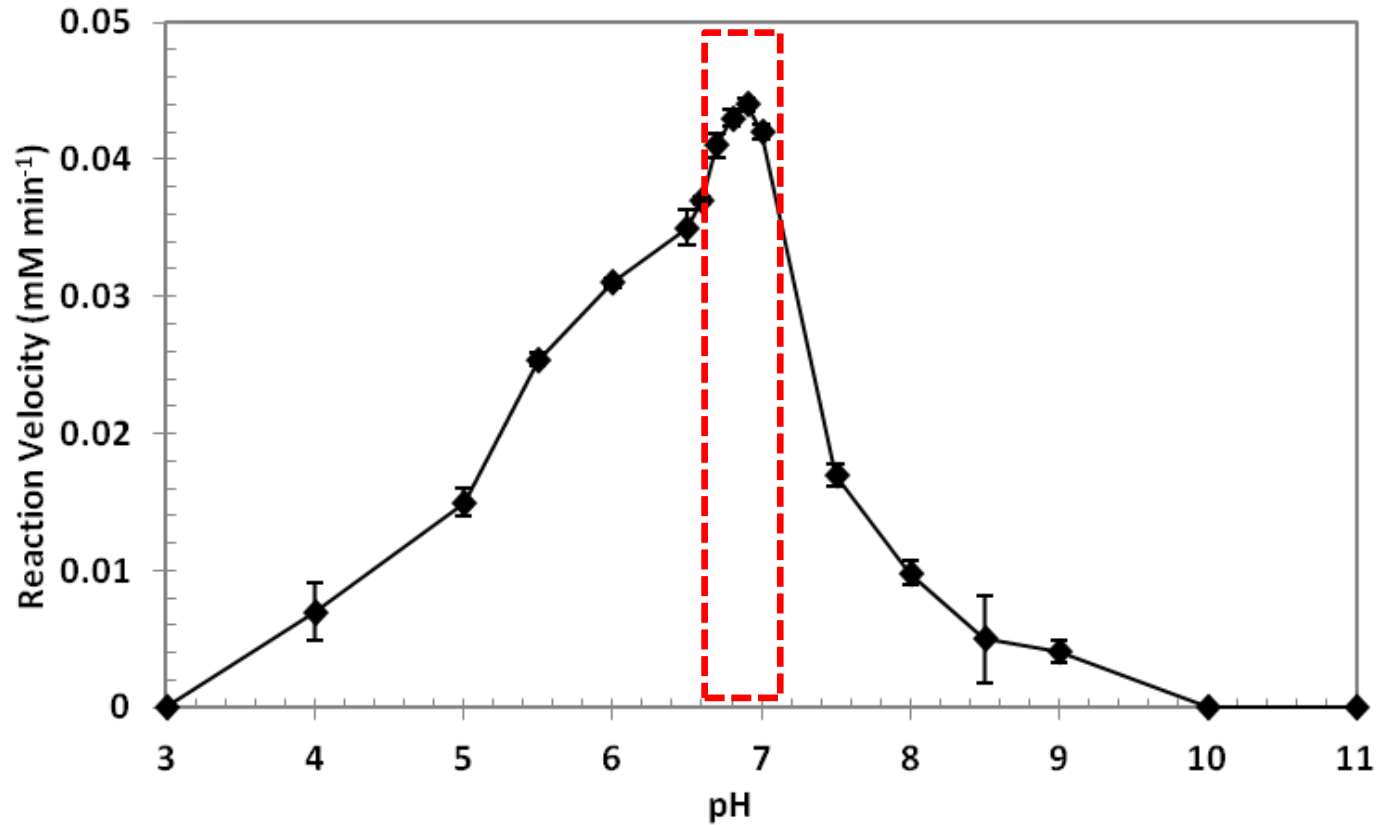
3M Petrifilms

Substrate  
uptake

# Background: How this works?

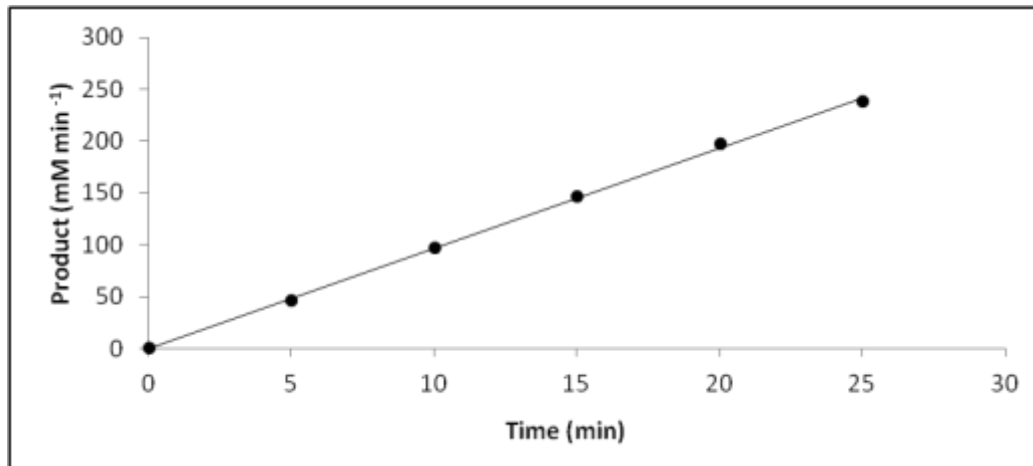
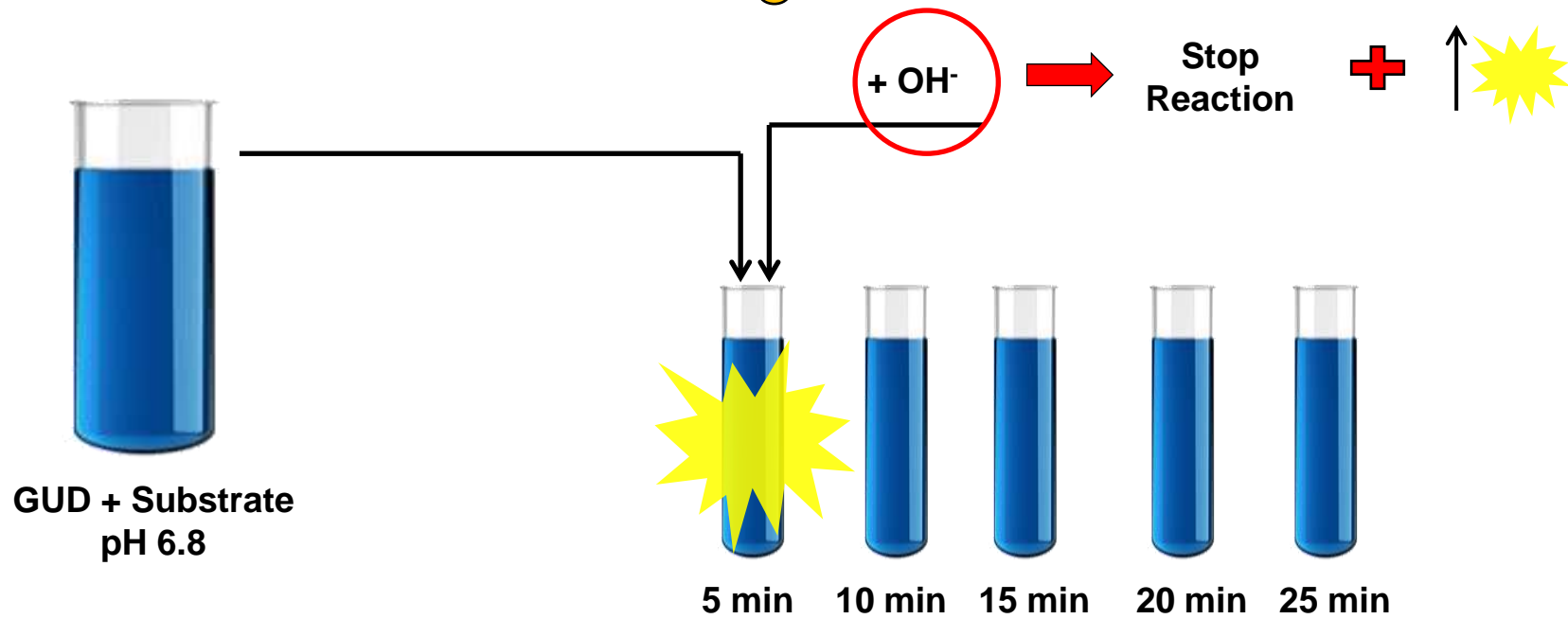


# Background: GUD pH optimum





# Background: Discontinuous Methods

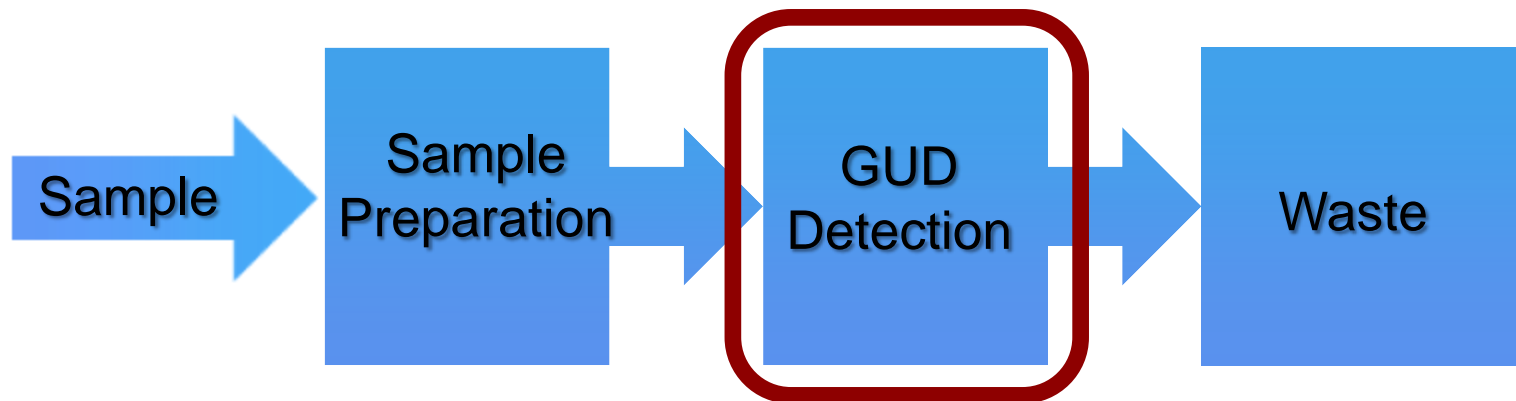


# Aim:

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□ Develop a continuous fluorometric method for the detection of GUD activity.

□ Develop a sensing platform for *E. Coli* detection in environmental waters.



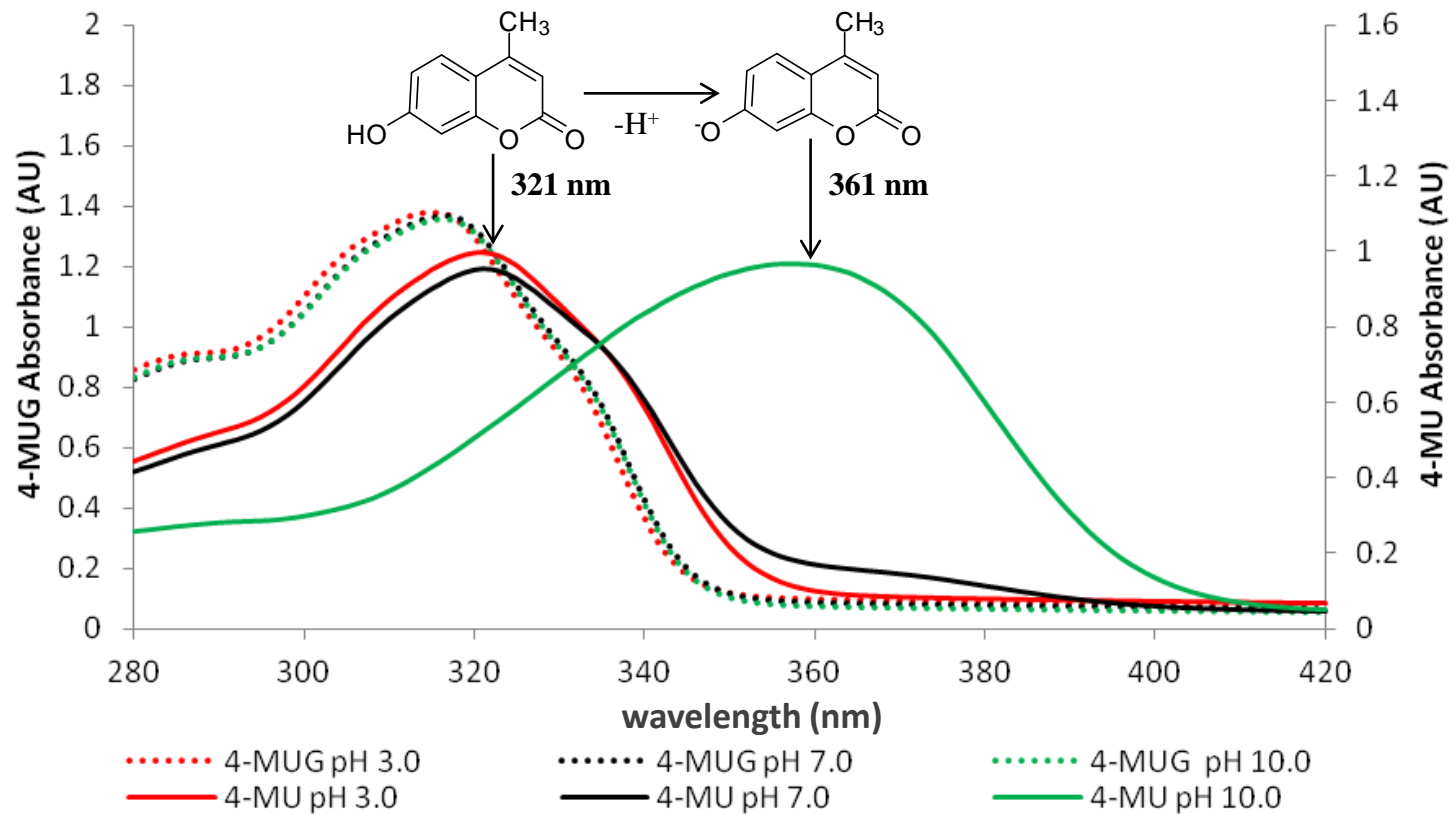
# Results

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- ❑ Comparison of 3 substrates for the measurement of GUD activity:
  - Ex/Em optimisation
  - GUD substrate kinetics
  
- ❑ 4-MUG/4-MU
- ❑ 3-CUG/3-CU
- ❑ 6-CMUG/6-CMU

# UV-VIS Characterization

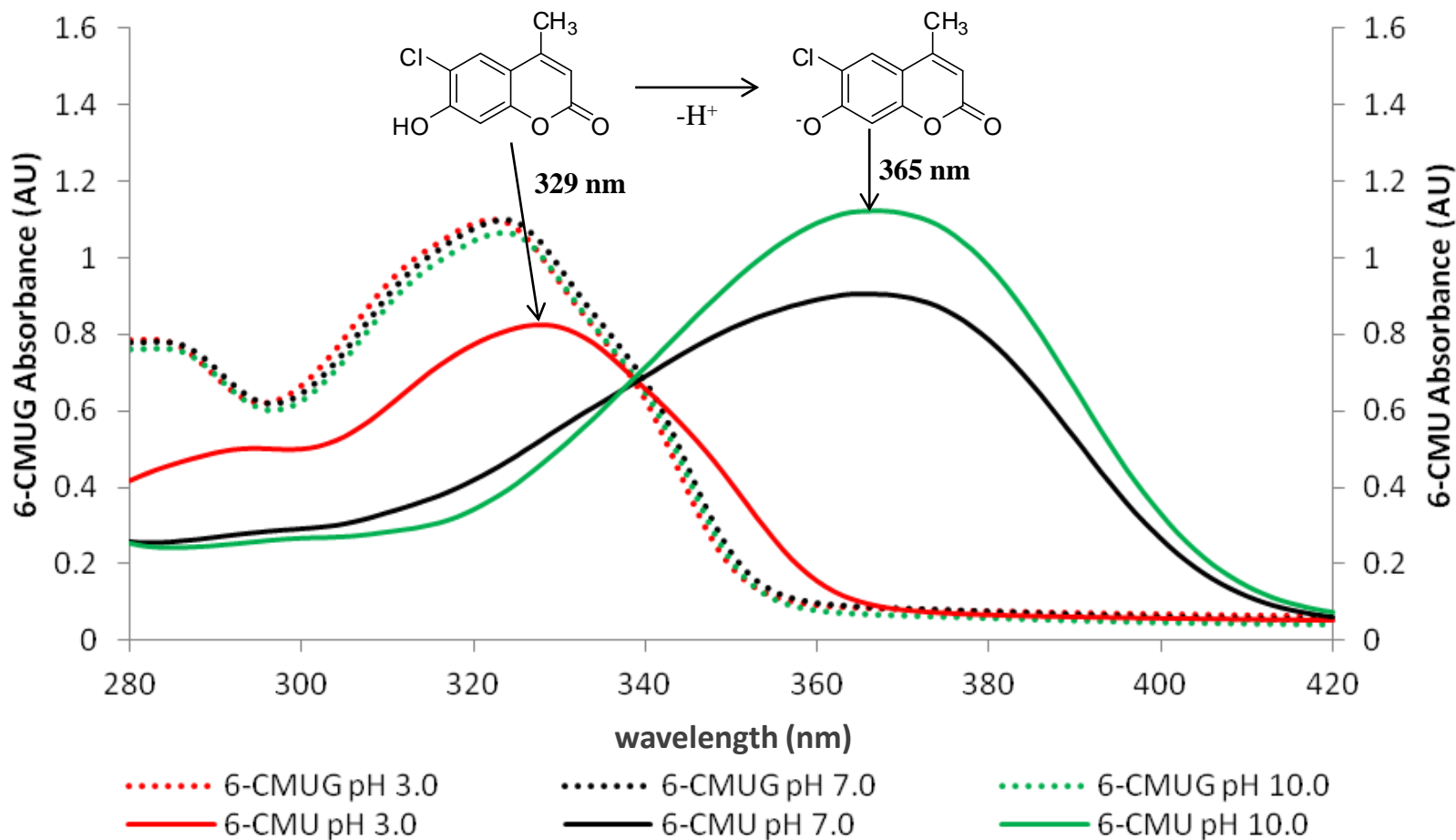
## □ 4-MUG / 4-MU



Absorption spectra of 100  $\mu\text{M}$  3-MUG and 50  $\mu\text{M}$  4-MU in acidic, neutral and alkaline conditions.

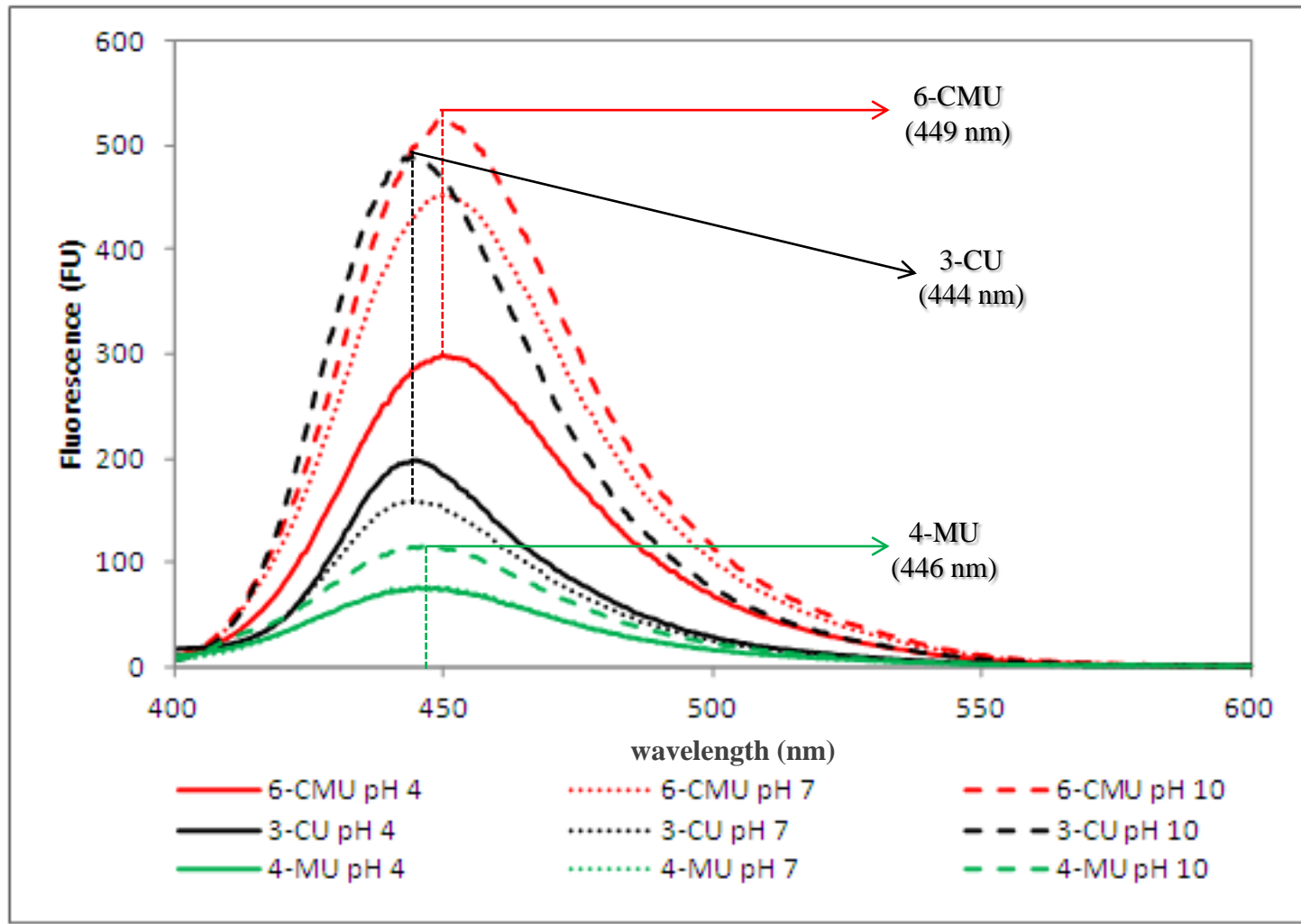
# Results – UV-VIS Characterization

## □ 6-CMUG / 6-CMU



Absorption spectra of 100  $\mu\text{M}$  6-CMUG and 50  $\mu\text{M}$  6-CMU in acidic, neutral and alkaline conditions.

# Ex/Em optimisation



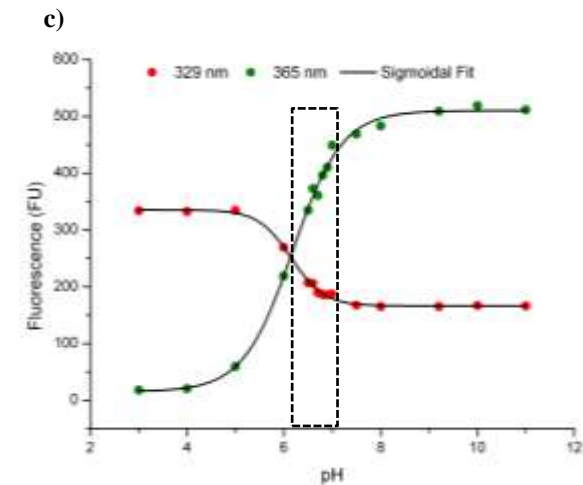
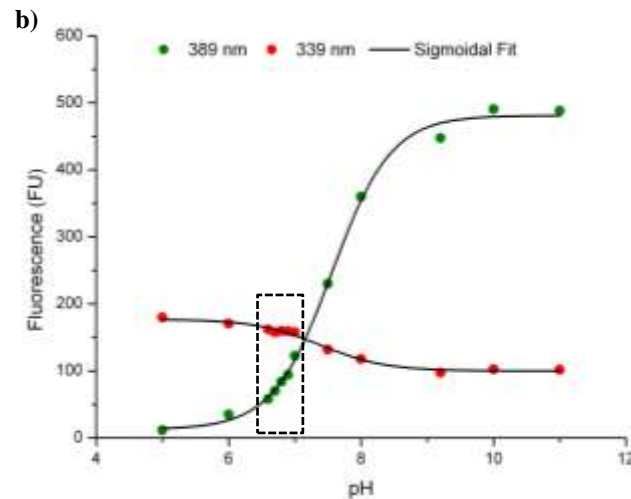
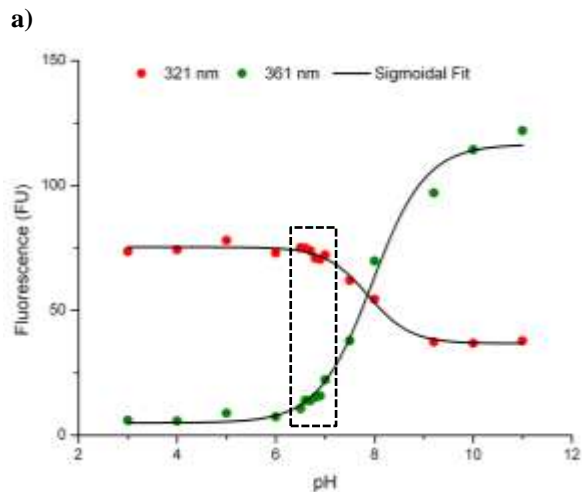
Emission spectra of 0.1  $\mu\text{M}$  4-MU, 3-CU and 6-CMU solutions in acidic, neutral and basic pH conditions. Each fluorophore was excited at the maximum excitation wavelength for the corresponding N and A<sup>-</sup> ground states.

# Ex/Em optimisation

4-MU pKa=  
 $7.86 \pm 0.6$

3-CU pKa=  
 $7.38 \pm 0.6$

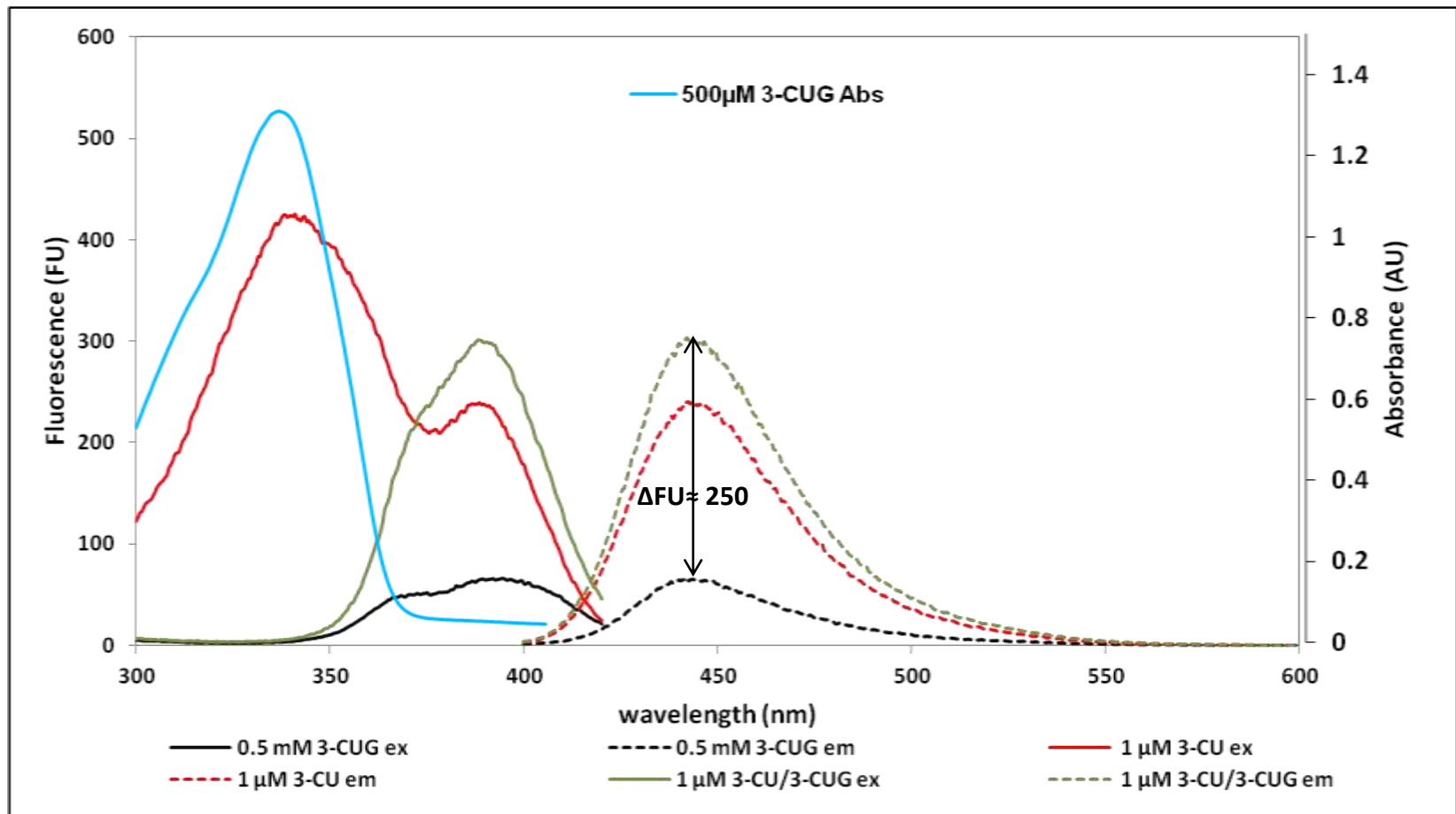
6-CMU pKa=  
 $6.18 \pm 0.3$



**Nonlinear regression fitting of the experimental data to Boltzman Sigmoidal model; Experimental data and model line for 4-MU (a), 3-CU (b) and 6-CMU (c). Excitation wavelengths are shown in the legend.**

# Ex/Em optimisation

## □ 3-CUG / 3-CU

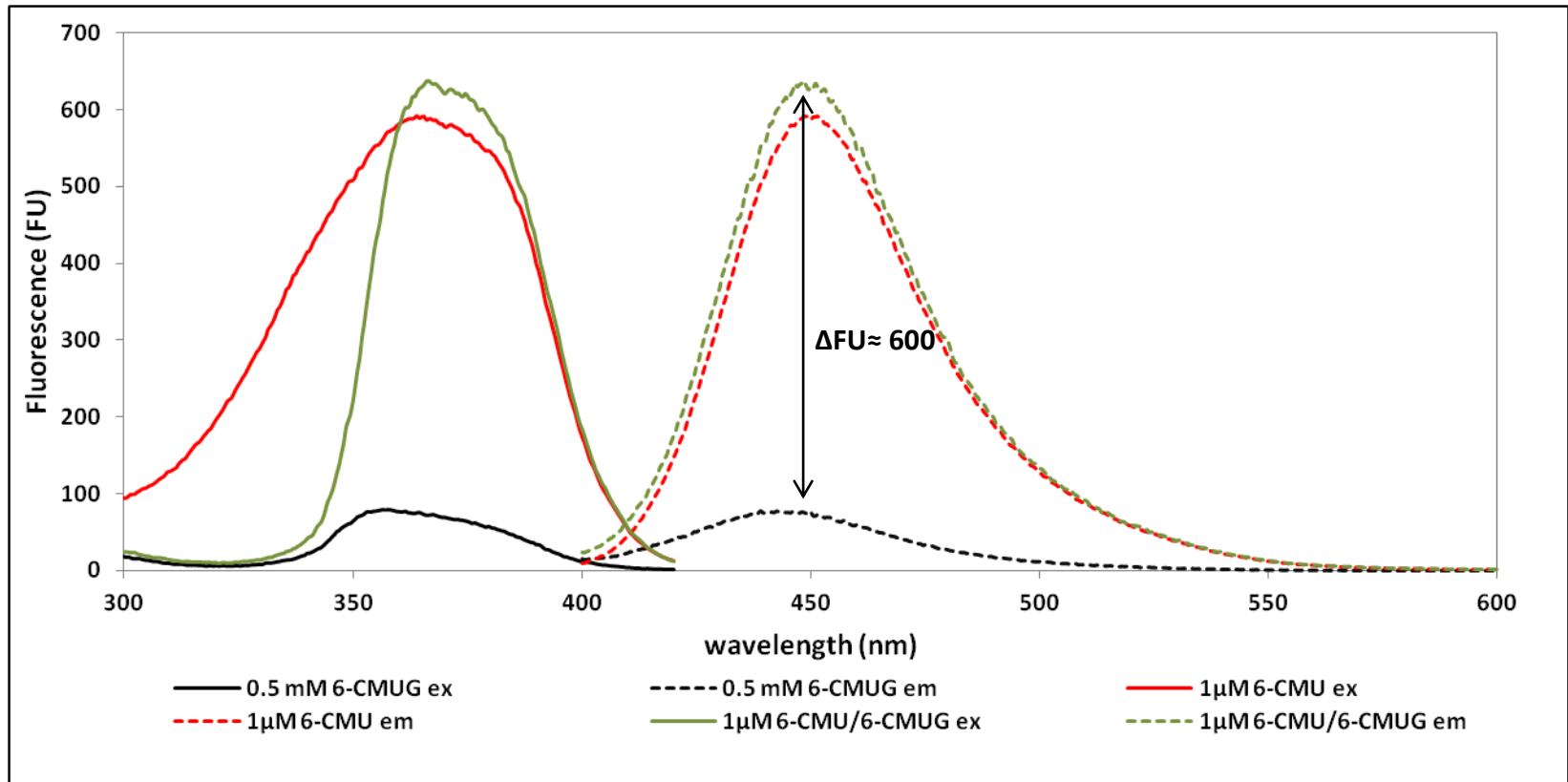


Excitation and emission spectra of 3-CU as influenced by the presence of 3-CUG in Phosphate buffer pH 6.8



# Ex/Em optimisation

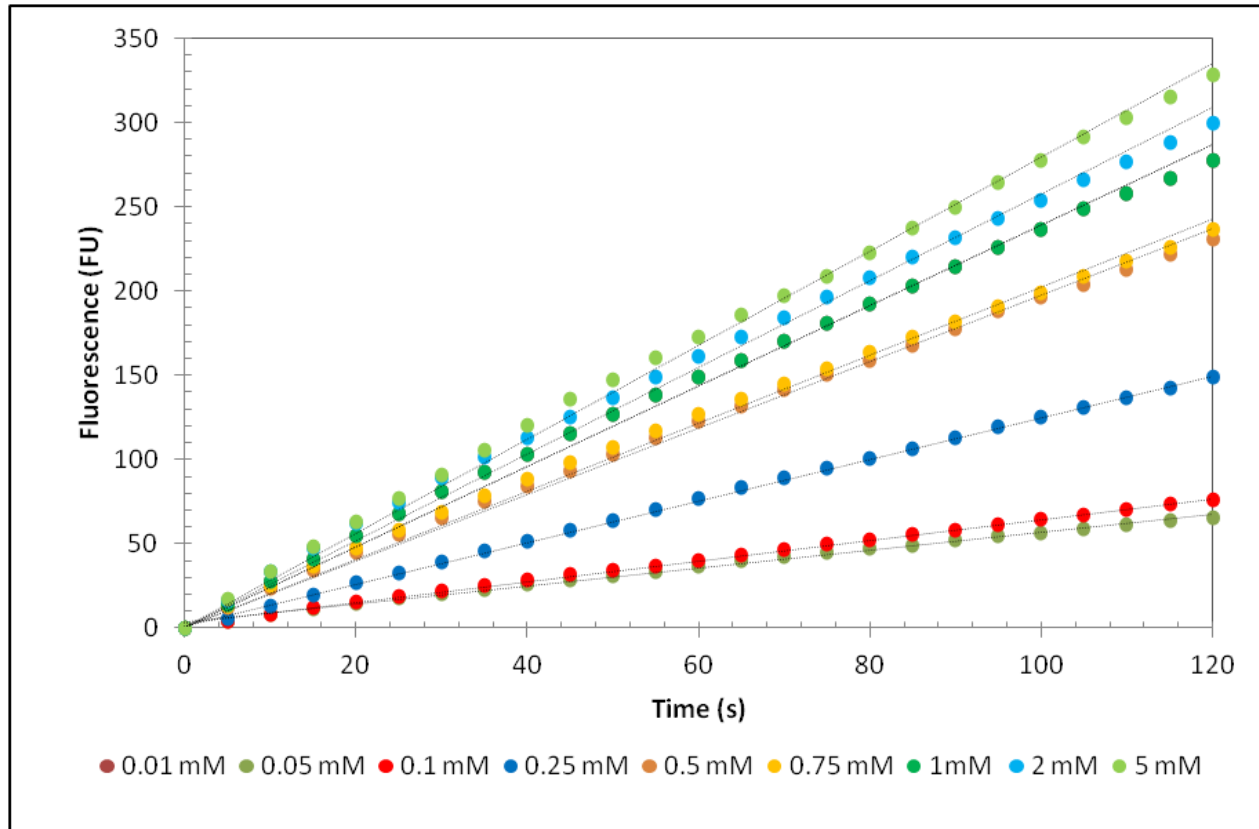
## □ 6-CMUG / 6-CMU



Excitation and emission spectra of 6-CMU as influenced by the presence of 6-CMUG in Phosphate buffer pH 6.8

# GUD-Substrate kinetics

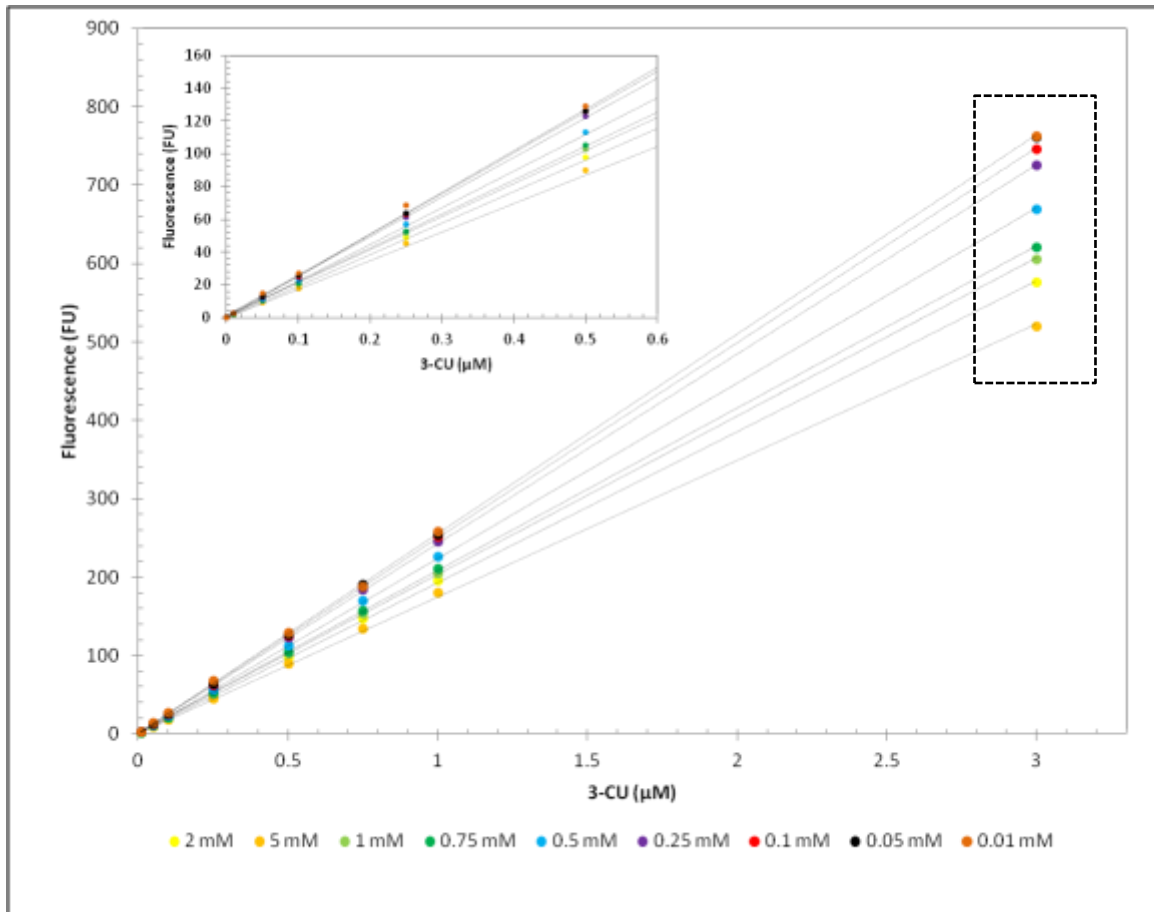
## □ Reaction progress curves



Reaction progress curves for GUD catalysed hydrolysis of different 3-CUG concentrations (shown in the legend);  $\lambda_{ex} = 389 \text{ nm}$ ,  $\lambda_{em} 444 \text{ nm}$ ; slit widths: 5 nm (ex), 2.5 nm (em); GUD was added at a concentration of 500 ng mL<sup>-1</sup>.

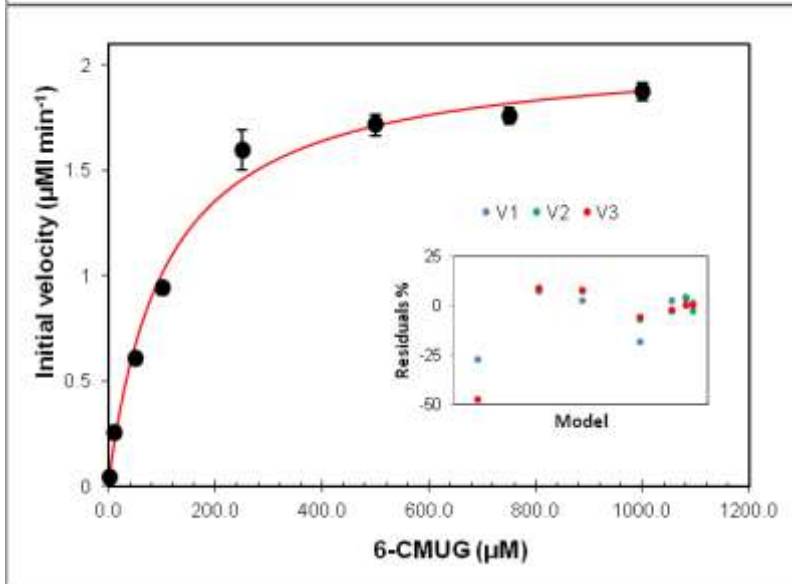
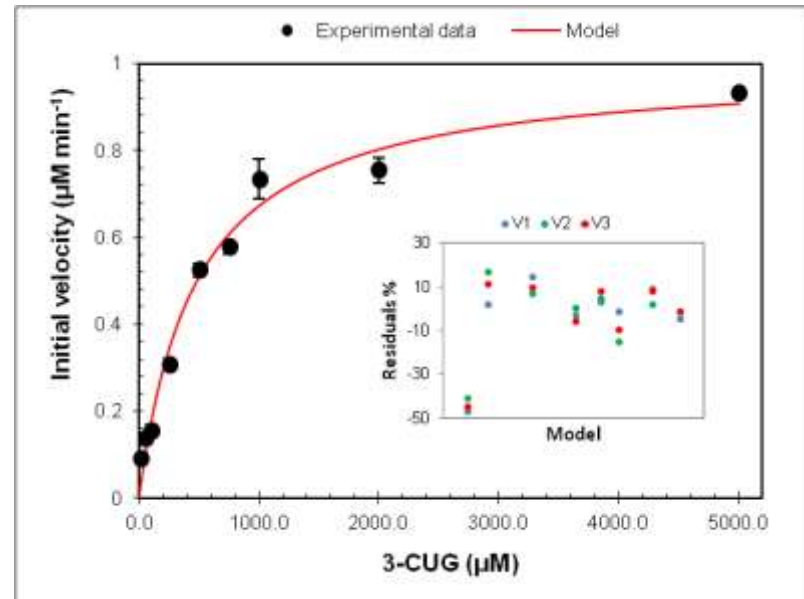
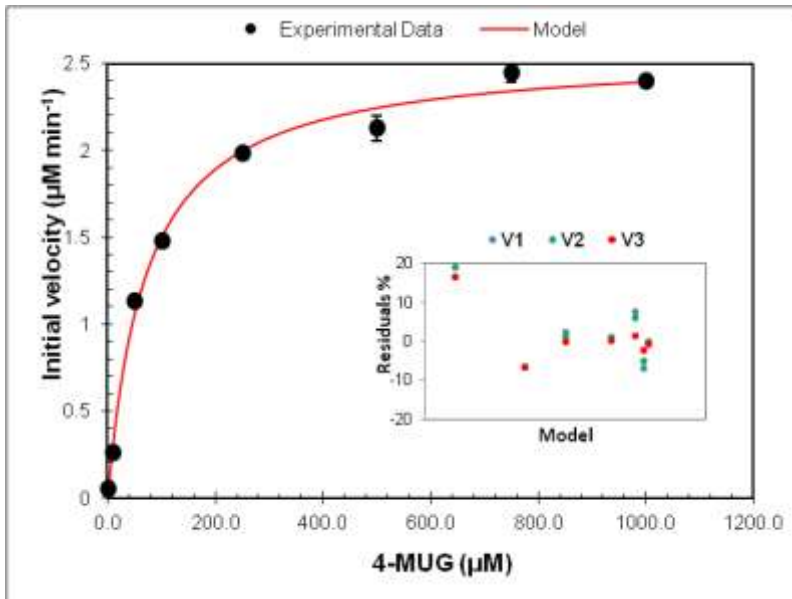
# GUD-Substrate kinetics

## □ Calibration



Calibration curves for 3-CU in the presence of different 3-CUG concentrations (shown in the legend);  $\lambda_{\text{ex}} = 389 \text{ nm}$ ,  $\lambda_{\text{em}} = 444 \text{ nm}$ ; slit widths: 5 nm (ex), 2.5 nm (em);

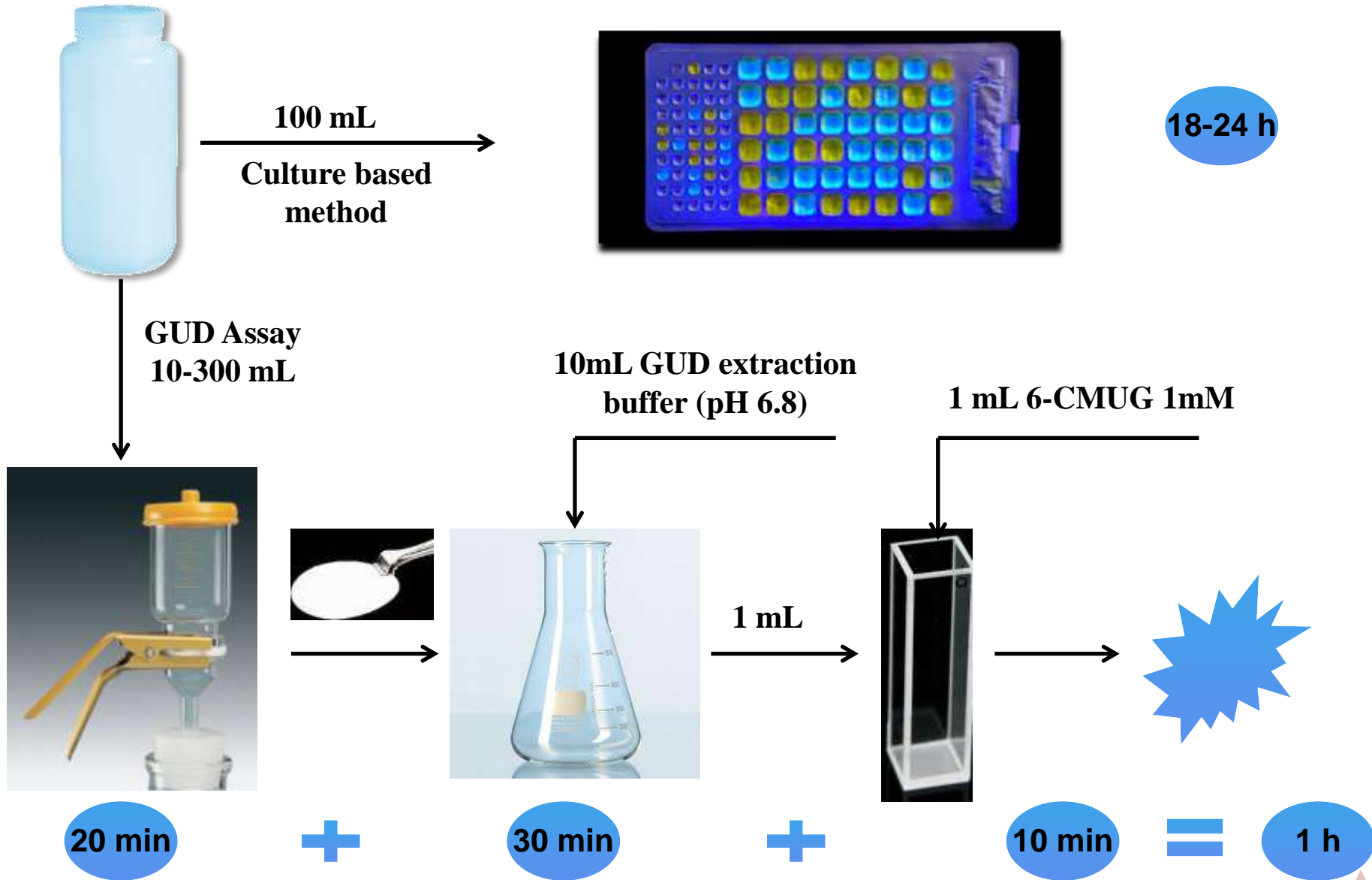
# GUD-Substrate kinetics



**Michaelis-Menten kinetic parameters (20 °C, pH 6.8)**

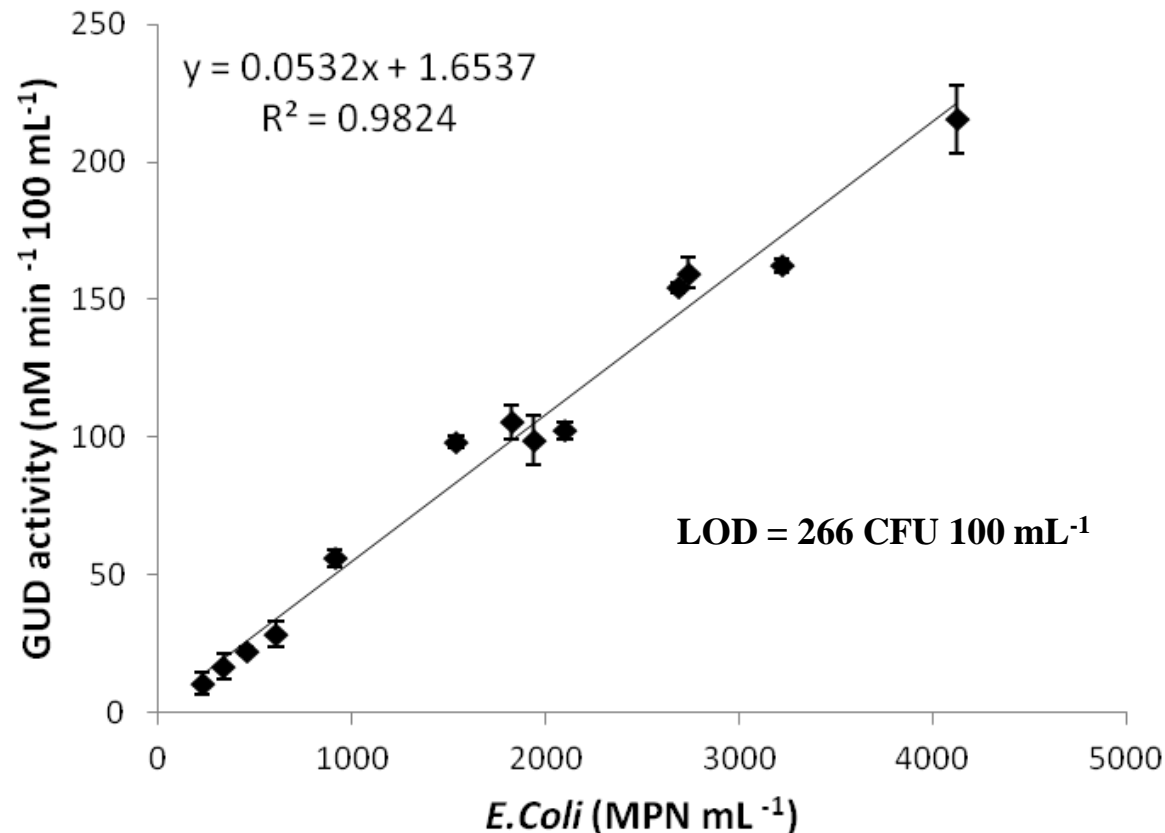
Substrate	$K_m$ ( $\mu\text{M}$ )	$V_{\max}$ ( $\mu\text{M min}^{-1}$ )	$V_{\max}/K_m$
4-MUG	70.82	2.56	0.031
3-CUG	479.28	0.99	0.002
6-CMUG	106.88	2.07	0.019

# Rapid *E. Coli* detection. Procedure



# Rapid *E. Coli* detection

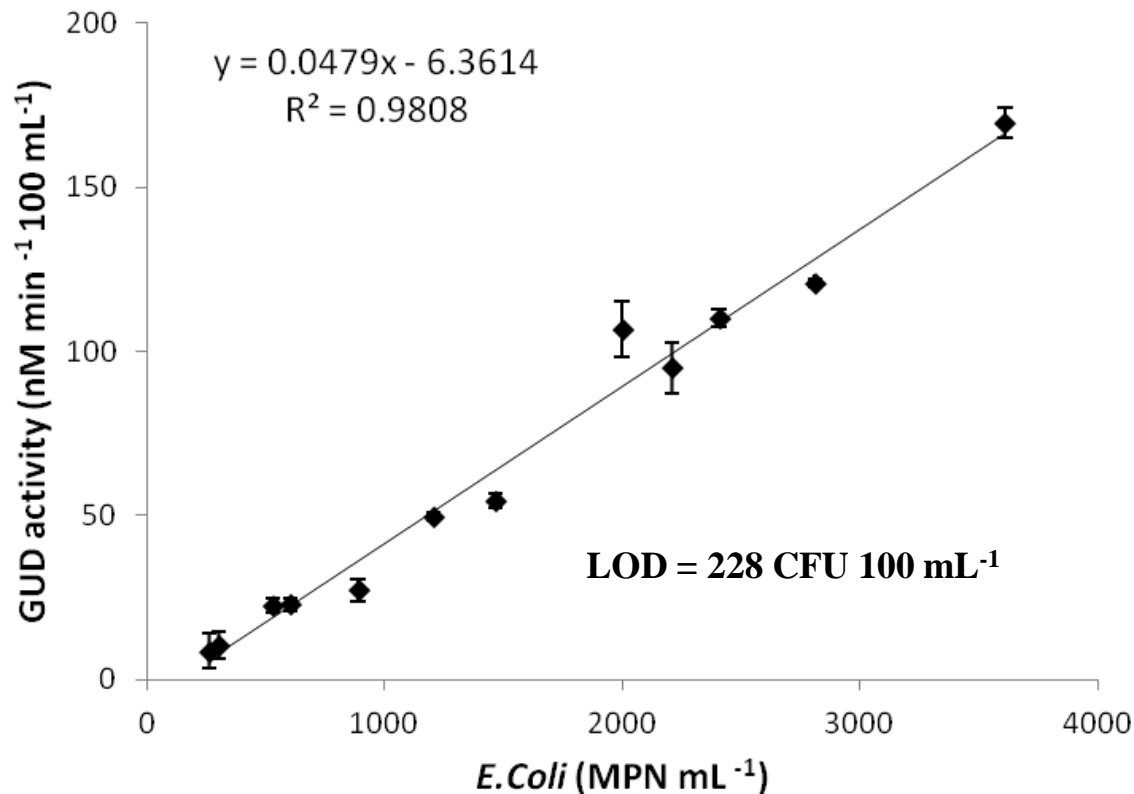
## □ Sea water



Linear regression between GUD activity and *E. Coli* concentrations in seawater. Error bars represent the SD of n=3 GUD activity measurements taken for each *E. Coli* concentration;

# Rapid *E. Coli* detection

## □ Fresh water



Linear regression between GUD activity and *E. Coli* concentrations in river water. Error bars represent the SD of n=3 GUD activity measurements taken for each *E. Coli* concentration;

# Conclusions/Impact:

- ❑ Continuous fluorometric method for the determination of GUD activity has been developed.
  
- ❑ Advantages
  - ✓ reagent consumption is minimised
  - ✓ possible to follow reaction kinetics only when a small amount of sample is available.
  - ✓ straightforward approach, prompt evaluation of kinetic data
  - ✓ minimal sample manipulation, experimental error reduced
  - ✓ potential for the implementation into an autonomous sensing platform
  
- ❑ The method was applied for the detection of *E. Coli* from environmental water samples and was successful in predicting *E. Coli* concentrations below the EU threshold for “excellent quality” , in 1h.
  
- ❑ Better management of bathing areas.



# Acknowledgements

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Prof. Fiona Regan

Brendan Heery



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Thank you all very much for your  
attention!

Questions?